

to control the microenvironment of the nucleation site and to manipulate near-surface gradients of concentrations of the crystallizing ions by patterning SAMs into rapidly and slowly nucleating regions.

The technique we report here gives us the ability to fabricate a large number of indistinguishable active nucleation regions, and to nucleate one crystal in each region. This should enable the study of fundamental aspects of the crystallization process by providing access to statistically significant numbers of nucleation events in highly controlled microenvironments. □

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Complete asymmetric induction of supramolecular chirality in a hydrogen-bonded assembly

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Chirality at the supramolecular level involves the non-symmetric arrangement of molecular components in a non-covalent assembly^{1,2}. Supramolecular chirality is abundant in biology, for example in the DNA double helix³, the triple helix of collagen⁴ and the α -helical coiled coil of myosin⁵. These structures are stabilized by inter-strand hydrogen bonds, and their handedness is deter-

mined by the configuration of chiral centres in the nucleotide or peptide backbone. Synthetic hydrogen-bonded assemblies have been reported that display supramolecular chirality in solution^{6–8} or in the solid state^{9–12}. Complete asymmetric induction of supramolecular chirality—the formation of assemblies of a single handedness—has been widely studied in polymeric superstructures^{13,14}. It has so far been achieved in inorganic metal-coordinated systems^{15–17}, but not in organic hydrogen-bonded assemblies^{18–20}. Here we describe the diastereoselective assembly of enantio-pure calix[4]arene dimelamines and 5,5-diethylbarbituric acid (DEB) into chiral hydrogen-bonded structures of one handedness. The system displays complete enantioselective self-resolution: the mixing of homomeric assemblies (composed of homochiral units) with opposite handedness does not lead to the formation of heteromeric assemblies. The non-covalent character of the chiral assemblies, the structural simplicity of the constituent building blocks and the ability to control the assembly process by means of peripheral chiral centres makes this system promising for the development of a wide range of homochiral supramolecular materials or enantioselective catalysts.

Previously we have shown by X-ray crystallography and ¹H NMR spectroscopy that assembly 1₃•DEB₆ exclusively forms as the staggered isomer A (D₃), which displays supramolecular chirality both in solution and in the solid state (see Fig. 1)^{21,22}. In the absence of any other source of chirality, this isomer exists as a racemic mixture of the *M*- and *P*-enantiomers.

We have now found that assembly of 3 equivalents of the chiral calix[4]arene dimelamines (*R,R*)-2 or (*S,S*)-3, having (*R*)- or (*S*)-1-phenylethylamine moieties, respectively, with 6 equivalents of DEB gives quantitatively the assemblies 2₃•DEB₆ (*M*-enantiomer) or 3₃•DEB₆ (*P*-enantiomer) (compounds 1–7 are shown in Fig. 1). The induction of helicity in both assemblies is complete (d.e. > 98%, where d.e. is diastereomeric excess) as judged from the single set of signals in the ¹H NMR spectra (CD₂Cl₂) (Fig. 2). None of the other possible diastereoisomeric assemblies (that is, (*P*)-2₃•DEB₆ and (*M*)-3₃•DEB₆) is present. Two-dimensional rotating frame Overhauser effect spectroscopy (ROESY) experiments correlate the absolute configuration of the substituents with the helicity: an (*R*)-substituent induces *M*-helicity, and an (*S*)-substituent induces *P*-helicity, in the assembly (Fig. 2d). We note that both assemblies are strongly active in circular dichroism (CD; $\Delta\epsilon_{\max} \approx 100 \text{ cm}^2 \text{ mmol}^{-1}$, where $\Delta\epsilon_{\max} = (\epsilon_L - \epsilon_R)_{286 \text{ nm}}$, Fig. 3a), while none of the individual components (*R,R*)-2 or (*S,S*)-3 show any significant CD activity ($\Delta\epsilon_{\max} < 8 \text{ cm}^2 \text{ mmol}^{-1}$). The observed CD is thus clearly a direct result of assembly formation and not an intrinsic property of the individual components.

Complete induction of chirality was also observed for the (*S*)-alanine- and (*R*)-naphthyldimelamine assemblies (*P*)-4₃•DEB₆ and (*M*)-5₃•DEB₆ (d.e. > 98% according to ¹H NMR spectroscopy; data not shown) and seems to be a general phenomenon for this type of assembly. Moreover, complete chiral induction is also observed with peripheral chiral centres in the cyanurate components. Combination of achiral dimelamine 6 with chiral cyanurates (*R*)- or (*S*)-MePhCYA or amino acid-derived cyanurates (*S*)-PheCYA, (*S*)-ValCYA, or (*S*)-LeuCYA (see Fig. 1 for nomenclature) leads in all cases to diastereoselective assembly of 6₃•CYA₆ with d.e. values > 98% according to ¹H NMR spectroscopy (data not shown). The CD spectra of the assemblies 2₃•DEB₆–5₃•DEB₆ and 6₃•CYA₆ all display bisignate curves with remarkably large amplitudes. The peripheral chromophores (benzyl, carbonyl, naphthyl) only affect the intensity of the Cotton effect (CE) at lower wavelengths, while the CD curves are virtually identical above 250 nm (Fig. 3). The observed Cotton effects seem to be largely the result of exciton coupling between chromophores present in the core of the assemblies. Comparison of the different CD spectra suggests that the sign of the CD curve is a good probe for the helicity of the assembly.

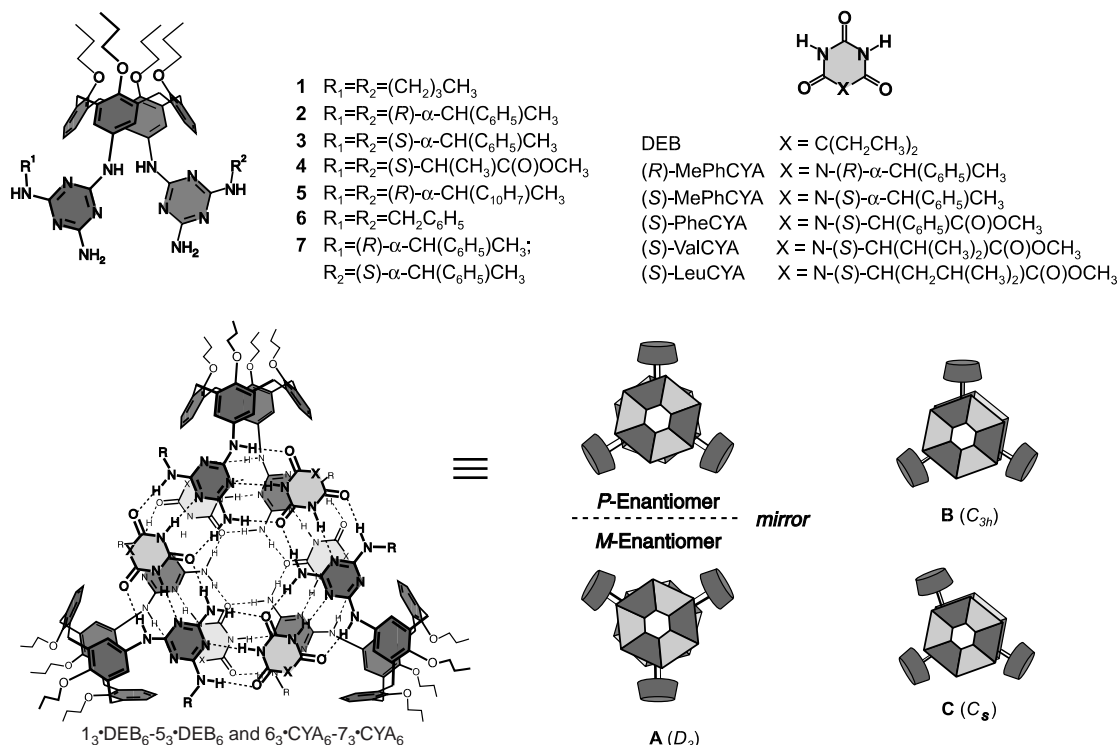


Figure 1 Schematic representations and molecular structure of the nine-component hydrogen bonded assemblies $1,3\text{-DEB}_6\text{-}5,3\text{-DEB}_6$, $6,3\text{-CYA}_6$ and $7,3\text{-CYA}_6$ with their individual components. The assemblies can exist in three isomeric forms: the chiral staggered isomer **A** (D_3 symmetry), and the achiral

eclipsed isomers **B** (C_{3h} symmetry) and **C** (C_s symmetry). The chiral isomer **A** can be present as a mixture of the *P*- and *M*-enantiomer. The assignment of the *M/P*-configuration is based on a clockwise (*P*) or anticlockwise (*M*) orientation of the two melamine units in the assembly²⁷.

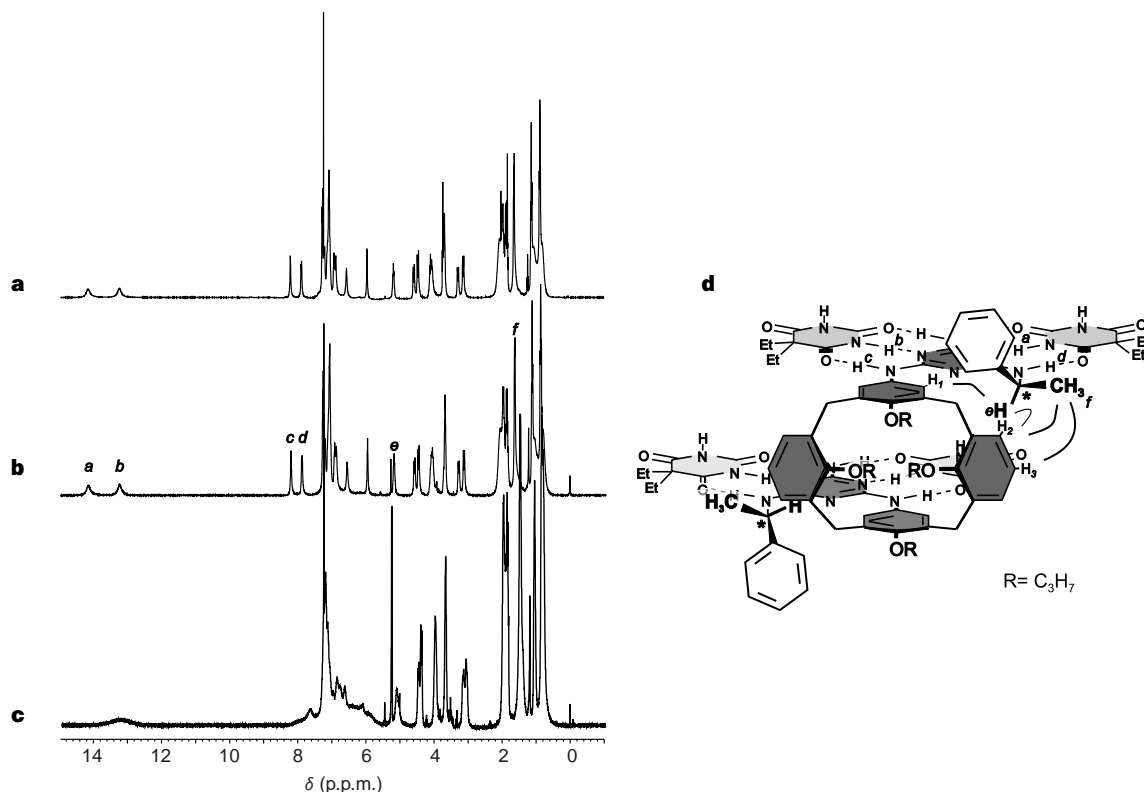


Figure 2 Characterization of chiral non-covalent assemblies (*M*)- $2,3\text{-DEB}_6$ and (*P*)- $3,3\text{-DEB}_6$ by 1H NMR spectroscopy. **a**, 400 MHz spectrum of enantiomerically pure assembly (*M*)- $2,3\text{-DEB}_6$; **b**, 400 MHz spectrum of enantiomerically pure assembly (*P*)- $3,3\text{-DEB}_6$; **c**, 400 MHz spectrum of the assembly of (*R,S*)-**7** and DEB (2 equiv.); all spectra were recorded in CD_2Cl_2 (1 mM) at 298 K relative to residual CH_2Cl_2 . **d**, 2D ROESY connectivities measured for assembly (*P*)- $3,3\text{-DEB}_6$ in toluene- d_8 . Medium connectivities were observed for the following protons: H_a

and H_b , H_c and H_d , and H_e , H_f and H_g , and H_h and H_i . These connectivities relate the configuration of the chiral substituent (*S*) with the helicity (*P*) of the assembly. Additional evidence for the formation of assemblies (*M*)- $2,3\text{-DEB}_6$ and (*P*)- $3,3\text{-DEB}_6$ was obtained by MALDI-TOF mass spectrometry using Ag^+ -labelling²⁸. Both assemblies exhibit a strong signal at m/z 4,358.3 (calculated for $C_{234}H_{288}N_{48}O_{30}^{107}Ag^+$, 4,360.2) corresponding to the monovalent Ag^+ -complexes.

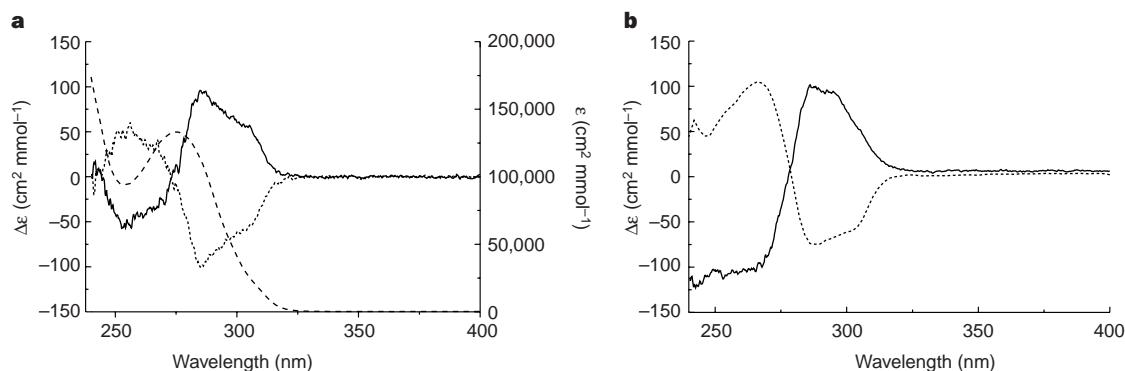


Figure 3 Characterization of assemblies $2_3\bullet\text{DEB}_6$ – $5_3\bullet\text{DEB}_6$ by CD and ultraviolet spectroscopy. **a**, CD spectra and ultraviolet spectrum (dashed line) of enantiomerically pure assemblies (M) - $2_3\bullet\text{DEB}_6$ (solid) and (P) - $3_3\bullet\text{DEB}_6$ (dotted); **b**, CD

spectra of enantiomerically pure assemblies (P) - $4_3\bullet\text{DEB}_6$ (dotted) and (M) - $5_3\bullet\text{DEB}_6$ (solid). Spectra were recorded in CH_2Cl_2 (1 mM) at 298 K.

The observed induction of supramolecular chirality is obviously related to the presence of six peripheral chiral centres in each assembly. The energy parameters for the chirality induction were determined using a theoretical model in which the difference in enthalpy between the M - and P -helix (ΔH_{total}) is regarded as the sum of six individual contributions of each chiral centre ($\Delta H_{\text{R/S}}$) (see Supplementary Information). Using this model, the values of ΔH_{total} and $\Delta H_{\text{R/S}}$ can be determined directly from the measured d.e. values via the relation: $\Delta H_{\text{total}} = 6\Delta H_{\text{R/S}} = -RT\ln[-(\text{d.e.}_{6\text{R}} + 100)/(\text{d.e.}_{6\text{R}} - 100)]$, where d.e._{6R} is the diastereomeric excess in an assembly with 6 (R)-substituents. For example, a d.e._{6R} > 98% gives $-\Delta H_{\text{total}}(298\text{ K}) > 11.4\text{ kJ mol}^{-1}$ and $-\Delta H_{\text{R/S}}(298\text{ K}) > 1.90\text{ kJ mol}^{-1}$. Cooperativity between the chiral centres is not included in the model, but will lead to the fact that $-\Delta H_{\text{R/S}} < -\Delta H_{\text{total}}/6$.

For a more accurate determination of $\Delta H_{\text{R/S}}$, we studied the heteromeric assemblies $2_{3-n}6_n\bullet\text{DEB}_6$ ($n = 1$ – 2), in which 1 or 2 chiral components (R,R -**2** are replaced by achiral components **6**. Determination of the corresponding d.e. values (d.e._{4R/2N} and d.e._{2R/4N})—which will be lower than d.e._{6R} as a result of the reduced number of chiral centres—will therefore allow a more accurate determination of $\Delta H_{\text{R/S}}$. The heteromeric assemblies $2_{3-n}6_n\bullet\text{DEB}_6$ ($n = 1$ – 2) cannot be obtained in pure form due to their dynamic character, but are formed spontaneously by mixing the homomeric assemblies (M) - $2_3\bullet\text{DEB}_6$ (Fig. 4a) and $6_3\bullet\text{DEB}_6$ (Fig. 4b), as indicated by multiple signals in the ^1H NMR spectrum (Fig. 4c). We have shown previously that mixtures of achiral assemblies have a

statistical composition²³. Titration of chiral assembly (M) - $2_3\bullet\text{DEB}_6$ with racemic assembly $6_3\bullet\text{DEB}_6$ followed by measurement of the CD spectra reveals a nonlinear relation between the chiroptical activity ($\Delta\epsilon_{286}$) of the mixture and the amount of added $6_3\bullet\text{DEB}_6$ (Fig. 4d). The chiroptical activity for each measurement is composed of three individual contributions: $\Delta\epsilon_{\text{total}} = (\text{d.e.}_{6\text{R}}[2_3\bullet\text{DEB}_6] + \text{d.e.}_{4\text{R}/2\text{N}}[2_26_1\bullet\text{DEB}_6] + \text{d.e.}_{2\text{R}/4\text{N}}[2_16_2\bullet\text{DEB}_6]) \times \Delta\epsilon(2_3\bullet\text{DEB}_6)$, assuming the same $\Delta\epsilon$ for each assembly $2_{3-n}6_n\bullet\text{DEB}_6$ ($n = 0$ – 3). Curve fitting (see Supplementary Information) using the theoretical model described above reveals the best fit for $-\Delta H_{\text{total}}(298\text{ K}) = 13.4\text{ kJ mol}^{-1}$ and $-\Delta H_{\text{R/S}}(298\text{ K}) = 2.23\text{ kJ mol}^{-1}$ (Fig. 4d), with corresponding d.e. values of 99.1% ($2_3\bullet\text{DEB}_6$), 94.7% ($2_26_1\bullet\text{DEB}_6$) and 71.7% ($2_16_2\bullet\text{DEB}_6$). The d.e._{6R} value of 99.1% for assembly (M) - $2_3\bullet\text{DEB}_6$ is in good agreement with that determined by ^1H NMR spectroscopy (see above). The chirality induction in the heteromeric assemblies $2_{3-n}6_n\bullet\text{DEB}_6$ ($n = 1$ – 2) resembles the ‘Sergeants and Soldiers’ principle (that is, the achiral units reinforce the induced chirality of the chiral units) as observed previously in polymeric²⁴ and liquid crystalline¹⁴ materials.

We also investigated mixtures of the chiral assemblies (M) - $2_3\bullet\text{DEB}_6$ and (P) - $3_3\bullet\text{DEB}_6$. The ^1H NMR spectrum of a 1:1 mixture of the assemblies is identical to that of the separate assemblies (Fig. 5a–c). Additional signals are not observed, which indicates that the heteromeric assemblies $2_{3-n}3_n\bullet\text{DEB}_6$ ($n = 1$ – 2) are not formed to a significant extent. Moreover, titration of (M) - $2_3\bullet\text{DEB}_6$ with (P) - $3_3\bullet\text{DEB}_6$ followed by CD shows a strictly

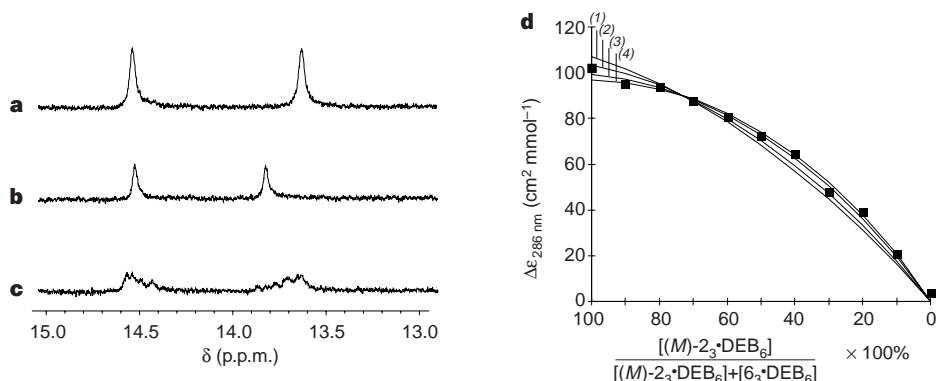


Figure 4 Characterization of mixed assemblies $2_{3-n}6_n\bullet\text{DEB}_6$ ($n = 0$ – 3) by ^1H NMR and CD spectroscopy. The figure shows the 13–15 p.p.m. region of the 300 MHz ^1H NMR spectrum of: **a**, enantiomerically pure assembly (M) - $2_3\bullet\text{DEB}_6$; **b**, racemic assembly $6_3\bullet\text{DEB}_6$; and **c**, a 1:1 mixture of (M) - $2_3\bullet\text{DEB}_6$ and $6_3\bullet\text{DEB}_6$. All spectra were recorded in toluene- d_8 (1 mM) at 298 K relative to residual

$\text{C}_6\text{D}_5\text{CHD}_2$. **d**, Plot of the CD intensity (filled squares, measured at 286 nm) versus the ratio $[(M)\text{-}2_3\bullet\text{DEB}_6]/[(M)\text{-}2_3\bullet\text{DEB}_6] + [6_3\bullet\text{DEB}_6] \times 100\%$. The lines represent the calculated curves for $\Delta H_{\text{total}} = -8.0(1)$, $-10.0(2)$, -13.4 (3, best fit) and -16.0 (4) kJ mol^{-1} .

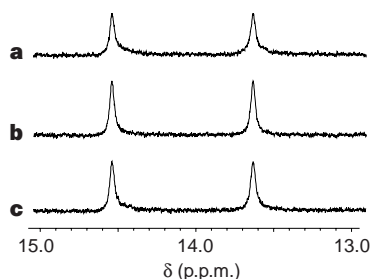
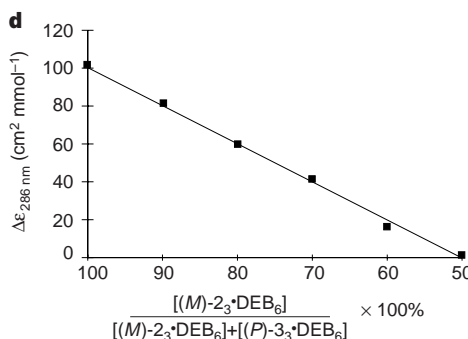


Figure 5 Characterization of mixed assemblies $2_3\text{-}n\text{-}3\text{-DEB}_6$ ($n = 0\text{--}3$) by ^1H NMR and CD spectroscopy. The figure shows the 13–15 p.p.m. region of the 300 MHz ^1H NMR spectrum of: **a**, enantiomerically pure assembly $(M)\text{-}2_3\text{-DEB}_6$; **b**, enantiomerically pure assembly $(P)\text{-}3_3\text{-DEB}_6$; and **c**, a 1:1 mixture of $(M)\text{-}2_3\text{-DEB}_6$ and



$(P)\text{-}3_3\text{-DEB}_6$. All spectra were recorded in toluene- d_8 (1 mM) at 298 K. **d**, Plot of the CD intensity (filled squares, measured at 286 nm) versus the ratio $[(M)\text{-}2_3\text{-DEB}_6]/[(M)\text{-}2_3\text{-DEB}_6] + [(P)\text{-}3_3\text{-DEB}_6] \times 100\%$. The solid line represents the calculated curve for $\Delta H_{\text{total}} = -15.0 \text{ kJ mol}^{-1}$.

linear decrease of the chiroptical activity (Fig. 5d). Curve fitting (see Supplementary Information) using the theoretical model reveals an improving fit for increasing $-\Delta H_{R/S}$ values. These results confirm the ^1H NMR experiment, and also indicate that the homomeric assemblies $(M)\text{-}2_3\text{-DEB}_6$ and $(P)\text{-}3_3\text{-DEB}_6$ are exclusively present in a mixture of the chiral building blocks $(R,R)\text{-}2$ and $(S,S)\text{-}3$ and DEB (3:3:6). In the case of statistical mixing, a 25:75 mixture of homomeric and heteromeric assemblies would be formed. The present system thus constitutes an example of complete enantioselective self-resolution in a dynamic hydrogen-bonded assembly in solution, a phenomenon so far only observed for hydrogen-bonded assemblies in the solid²⁵ or liquid-crystalline²⁶ state.

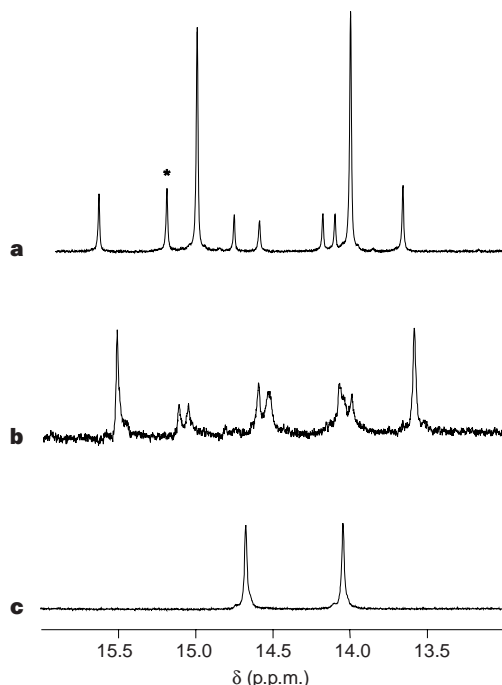


Figure 6 Isomeric distribution of hydrogen-bonded assemblies from **1**, $(R,R)\text{-}2$, and $(R,S)\text{-}7$ with HexCYA as determined by ^1H NMR spectroscopy. 300 MHz ^1H NMR spectrum (13–16 p.p.m.) of: **a**, assembly 1_3-HexCYA_6 (mixture of isomers **A**–**C**; the signal denoted with an asterisk represents two chemically different protons); **b**, *meso*-assembly 7_3-HexCYA_6 (mixture of isomers **B** and **C**); and **c**, enantiomerically pure assembly $(M)\text{-}2_3\text{-HexCYA}_6$ (exclusive formation of isomer **A**). Additional evidence for the formation of assemblies $(M)\text{-}2_3\text{-HexCYA}_6$ and 7_3-HexCYA_6 was obtained by MALDI-TOF mass spectrometry using Ag^+ -labelling²⁸. Both assemblies exhibit a strong signal at m/z 4,536.4 (calculated for $\text{C}_{240}\text{H}_{308}\text{N}_{54}\text{O}_{30}\text{Ag}^+$, 4,535) corresponding to the monovalent Ag^+ -complexes.

Chiral information in the individual components can also be used to control the conformation of the assembly. As we have shown above, both $(R,R)\text{-}2$ and $(S,S)\text{-}3$, which have two chiral centres of identical configuration, preferentially assemble with DEB as the staggered isomer **A** (Fig. 1). In sharp contrast to this, dimelamine $(R,S)\text{-}7$, which has two chiral centres of opposite configuration, is not able to form isomer **A** in the presence of 2 equivalents of DEB. In this case, non-defined oligomeric assemblies are formed that show broad signals in the ^1H NMR spectrum (Fig. 2c). More informative are the results obtained from assembly studies of **1**, $(R,R)\text{-}2$, and $(R,S)\text{-}7$ with HexCYA. Assembly 1_3-HexCYA_6 exists as a mixture of isomer **A** (D_3) and the two achiral eclipsed isomers **B** (C_{3h}) and **C** (C_s) (Fig. 1). The ^1H NMR spectrum exhibits 10 different signals in the 14–16 p.p.m. region; that is, two signals each for **A** and **B**, and six signals for **C** (Fig. 6a). However, for $(R,R)\text{-}2$ and $(R,S)\text{-}7$ the isomeric distribution of the assembly with HexCYA is changed dramatically. Assembly of 3 equivalents of $(R,S)\text{-}7$ with 6 equivalents of HexCYA gives a mixture of the two eclipsed isomers **B** and **C**. The ^1H NMR spectrum shows only eight signals in the 14–16 p.p.m. region, and isomer **A** is not present (Fig. 6b). In contrast to this, assembly of 3 equivalents of $(R,R)\text{-}2$ and 6 equivalents of HexCYA gives isomer **A** as the only product. Not a trace of the isomers **B** and **C** is found in the ^1H NMR spectrum; there are only two signals in the 14–16 p.p.m. region (Fig. 6c). Assembly 2_3-HexCYA_6 exhibits a CD spectrum characteristic of assemblies of the isomer **A** type. These experiments again illustrate the decisive role that the peripheral chiral centres play in the assembly process. □

Methods

Hydrogen-bonded assemblies were prepared by suspending calix[4]arene dimelamines **1**–**7** and either DEB or CYA in a 1:2 molar ratio in toluene or CH_2Cl_2 . Clear solutions were obtained after stirring the mixtures for 15 min at room temperature. Occasionally sonication or heating was required to dissolve the components. For the CD-titration experiments 1 mM solutions of the homomeric assemblies in CH_2Cl_2 were mixed in ratios of 10:1 to 1:10 for $(M)\text{-}2_3\text{-DEB}_6$ and 6_3-DEB_6 and 10:1 to 5:5 for $(M)\text{-}2_3\text{-DEB}_6$ and $(P)\text{-}3_3\text{-DEB}_6$. The CD spectra were recorded 15 min after mixing the separate solutions.

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Supplementary information is available on Nature's World-Wide Web site (<http://www.nature.com>) or as paper copy from the London editorial office of Nature.

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Variability of inorganic and organic phosphorus turnover rates in the coastal ocean

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Phosphorus is an essential nutrient in pelagic marine ecosystems. Phosphorus cycling in the upper ocean is, however, poorly understood, and few studies have directly investigated the biological utilization of this essential element^{1–4}. Here, we have determined *in situ* phosphorus-turnover rates in a coastal marine environment by measuring the activities of two cosmogenic radionuclides (^{32}P and ^{33}P , with half lives of 14.3 and 25.3 days, respectively) in dissolved inorganic, dissolved organic and total particulate phosphorus pools over a seasonal cycle. Phosphorus turnover rates within dissolved and particulate pools are rapid and vary over seasonal timescales, suggesting that low phosphorus concentrations can support relatively high primary production. Furthermore, picoplankton, such as bacteria, appear preferentially to

utilize certain dissolved organic phosphorus compounds to obtain other associated nutrients, such as carbon and nitrogen. It seems that the significance of the roles of both dissolved inorganic and organic phosphorus in supporting primary production—and, hence, CO_2 uptake and particulate organic carbon export—has been hitherto underestimated.

The radionuclides ^{32}P and ^{33}P are produced primarily by cosmic ray interactions with atmospheric argon and enter the oceans predominantly in rain^{5–7}. If the ratio of $^{33}\text{P}/^{32}\text{P}$ in rain is known, then one can determine the relative ‘age’ of cosmogenic phosphorus, P, by measuring the $^{33}\text{P}/^{32}\text{P}$ ratio in various biological pools. High $^{33}\text{P}/^{32}\text{P}$ ratios indicate an older P pool. The inventories of ^{32}P and ^{33}P in the ocean are quite low, ranging from just tens to hundreds of disintegrations per minute per square metre (d.p.m. m^{-2})^{1,8–10}. Thus, ^{32}P and ^{33}P measurements require several thousand litres of sea water and extensive purification from other β -emitters. Previous investigations which sought to utilize these isotopes were hampered by a lack of known input fluxes, possible contamination and the inability to measure the low-energy β -emitter ^{33}P , especially in coastal environments with high P concentrations^{1,8–10}. This study is, to our knowledge, the first to constrain the ^{32}P and ^{33}P input flux⁶ and to simultaneously measure both these isotopes in various dissolved inorganic, organic and particulate pools.

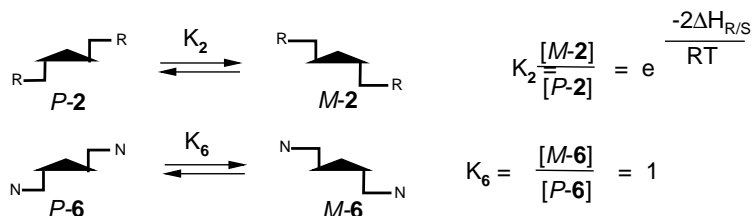
Sampling was conducted in Wilkinson basin in the Gulf of Maine ($42^\circ 29.41' \text{N}$, $69^\circ 45.02' \text{W}$) during four cruises in March, April, July and August 1997. This highly productive region supports one of the largest fisheries in North America¹¹. Surface and deep particulate and total dissolved phosphorus (TDP) samples were collected by passing 4,000–6,000 l of sea water sequentially through a series of 10, 1.0 and 0.2 μm cartridge prefilters followed by cartridges packed with iron-impregnated polypropylene filters. These filters have been demonstrated to collect TDP with close to 100% efficiency⁷. Separate surface samples were collected for soluble reactive phosphorus (SRP), as defined by the molybdenum blue method¹², using acrilan filters and the technique developed by Lee *et al.*⁹. Plankton tows (nominally $>102 \mu\text{m}$) were collected from various depths and sieved through a 335- μm screen to collect different size classes. All samples were extensively purified to remove all other β -emitting radionuclides and counted using low-level liquid scintillation⁶. In March and April, deep-water P samples were taken just above the base of the mixed layer (defined by a change in density $\geq 0.125 \text{ kg m}^{-3}$), whereas in July and August, deep-water samples were taken below both the mixed layer and the deep chlorophyll maximum.

The ratio of $^{33}\text{P}/^{32}\text{P}$ measured in rain at Portsmouth, NH and Woods Hole, MA was flux weighted over a 35 ± 3 -day period before the April, July and August cruises and over a 14-day period before the March cruise. $^{33}\text{P}/^{32}\text{P}$ ratios averaged 0.82 ± 0.07 (Fig. 1, Table 1). Thus, any ratio higher than this value must be due to radioactive decay. A non-continuous model can be used to determine the relative age of phosphorus in any particular reservoir: $\tau_p = [\ln(R_p/R_s)]/(\lambda_{32} - \lambda_{33})$ where τ_p is the age of phosphorus in the product material, R_p and R_s are the $^{33}\text{P}/^{32}\text{P}$ ratio found in the product and source material, respectively, and λ_{32} and λ_{33} are the radioactive decay constants⁵. Using this model, phosphorus ages are resolved on timescales ranging from 1 to 100 days. In general, age estimate errors will increase with increasing $^{33}\text{P}/^{32}\text{P}$ ratios as P activities decrease over time.

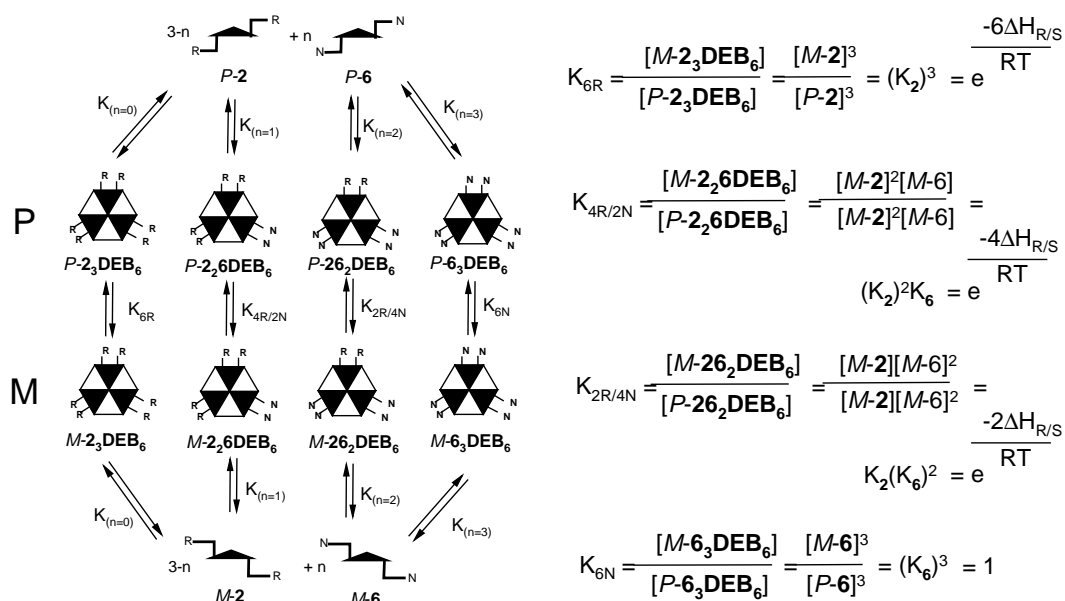
In March, April and July, ratios of $^{33}\text{P}/^{32}\text{P}$ in particulate matter were similar between surface and deep waters, indicating rapid transport of sinking particulate material from the euphotic zone to depth (Fig. 1). In August, the activities in all particulate samples retrieved from deep waters were below detection, indicating that the source of sinking particulates had decreased. $^{33}\text{P}/^{32}\text{P}$ ratios in total dissolved and small particulate ($<102 \mu\text{m}$) surface pools measured during March, April and August were similar to those found in rain.

Supplementary Information 2.1:
Model for Mixing Chiral Assembly 2_3-DEB_6
with Achiral Assembly 6_3-DEB_6

1) Define two different conformations *M* en *P* for separate building blocks **2** and **6**.



2) Incorporate *M/P*-conformations without change in energy into the assemblies.



N.B.: cooperativity between the chiral centres is not included in the model !!!

3) Curve fitting procedure (see attachment).

4) Calculate d.e._{6R}, d.e._{4R/2N} and d.e._{2R/4N} from optimized $\Delta H_{R/S}$ values from curve fitting.

$$d.e._{6R} = 100 \times \frac{(K_{6R} - 1)}{(K_{6R} + 1)}$$

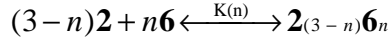
$$d.e._{4R/2N} = 100 \times \frac{(K_{4R/2N} - 1)}{(K_{4R/2N} + 1)}$$

$$d.e._{2R/4N} = 100 \times \frac{(K_{2R/4N} - 1)}{(K_{2R/4N} + 1)}$$

Supplementary Information 2.2:

Curve Fitting Procedure for Mixed Assemblies $2_{3-n}6_n \bullet \text{DEB}_6$ ($n=0-3$).

The equilibrium between assemblies and individual building blocks is described by



$$K(n) = \frac{[2_{(3-n)}6_n]}{[2]^{(3-n)}[6]^n}$$

$3K_{(n=0)}=K_{(n=1)}=K_{(n=2)}=3K_{(n=3)}$, as a result of the three possibilities of forming an assembly with composition **226** (**226**, **262**, and **622**) and **266** (**266**, **626**, and **662**). It is assumed that free components **2** and **6** are not present and therefore $K_{(n)}$ is assigned a value of $1E10$.

Assemblies with *P*-chirality are only formed from combinations of building blocks with *P*-conformation, *e.g.* *P*-**2** and *P*-**6**. The enthalpy of formation is identical for all assemblies (homomeric and heteromeric).

Based on these assumptions the mass balances for $[2_{\text{tot}}]$ and $[6_{\text{tot}}]$ are :

$$[2]_{\text{tot}} = 3[P\text{-}2_3 \bullet \text{DEB}_6] + 2[P\text{-}2_2 6 \bullet \text{DEB}_6] + [P\text{-}2 6_2 \bullet \text{DEB}_6] + 3[M\text{-}2_3 \bullet \text{DEB}_6] + 2[M\text{-}2_2 6 \bullet \text{DEB}_6] + [M\text{-}2 6_2 \bullet \text{DEB}_6].$$

$$[2]_{\text{tot}} = 3K_{(n=3)}[P\text{-}2]^3 + 2K_{(n=2)}[P\text{-}2]^2[P\text{-}6] + K_{(n=1)}[P\text{-}2][P\text{-}6]^2 + 3K_{(n=3)}[M\text{-}2]^3 + 2K_{(n=2)}[M\text{-}2]^2[M\text{-}6] + K_{(n=1)}[M\text{-}2][M\text{-}6]^2.$$

$$[2]_{\text{tot}} = 3K_{(n=3)}[P\text{-}2]^3 + 6K_{(n=3)}[P\text{-}2]^2[P\text{-}6] + 3K_{(n=3)}[P\text{-}2][P\text{-}6]^2 + 3K_{(n=3)}[M\text{-}2]^3 + 6K_{(n=3)}[M\text{-}2]^2[M\text{-}6] + 3K_{(n=3)}[M\text{-}2][M\text{-}6]^2.$$

$$[2]_{\text{tot}}/3K_{(n=3)} = [P\text{-}2]^3 + 2[P\text{-}2]^2[P\text{-}6] + [P\text{-}2][P\text{-}6]^2 + [M\text{-}2]^3 + 2[M\text{-}2]^2[M\text{-}6] + [M\text{-}2][M\text{-}6]^2.$$

$$[M\text{-}2] = K_2 \times [P\text{-}2] \text{ and } [M\text{-}6] = K_6 \times [P\text{-}6] \text{ gives:}$$

$$[2]_{\text{tot}}/3K_{(n=3)} = [P\text{-}2]^3(1+K_2^3) + 2[P\text{-}2]^2[P\text{-}6](1+K_2^2K_6) + [P\text{-}2][P\text{-}6]^2(1+K_2K_6^2).$$

With $K_6 = 1$:

$$= [P\text{-}2]^3(1+K_2^3) + 2[P\text{-}2]^2[P\text{-}6](1+K_2^2) + [P\text{-}2][P\text{-}6]^2(1+K_2).$$

This equation expresses $[P\text{-}6]$ as a function of K_2 and $[P\text{-}2]$.

$$[6]_{\text{tot}} = 3[P\text{-}6_3 \bullet \text{DEB}_6] + 2[P\text{-}2 6_2 \bullet \text{DEB}_6] + [P\text{-}2_2 6 \bullet \text{DEB}_6] + 3[M\text{-}6_3 \bullet \text{DEB}_6] + 2[M\text{-}2 6_2 \bullet \text{DEB}_6] + [M\text{-}2_2 6 \bullet \text{DEB}_6].$$

$$[6]_{\text{tot}} = 3K_{(n=0)}[P\text{-}6]^3 + 2K_{(n=1)}[P\text{-}2][P\text{-}6]^2 + K_{(n=2)}[P\text{-}2]^2[P\text{-}6] +$$

$$3K_{(n=0)}[M-6]^3 + 2K_{(n=1)}[M-2][M-6]^2 + K_{(n=2)}[M-2]^2[M-6].$$

$$[6]_{\text{tot}} = 3K_{(n=0)}[P-6]^3 + 6K_{(n=0)}[P-2][P-6]^2 + 3K_{(n=0)}[P-2]^2[P-6] + 3K_{(n=0)}[M-6]^3 + 6K_{(n=0)}[M-2][M-6]^2 + 3K_{(n=0)}[M-2]^2[M-6].$$

$$[6]_{\text{tot}}/3K_{(n=0)} = [P-6]^3 + 2[P-2][P-6]^2 + [P-2]^2[P-6] + [M-6]^3 + 2[M-2][M-6]^2 + [M-2]^2[M-6].$$

$[M-2] = K_2 \times [P-2]$ and
 $[M-6] = K_6 \times [P-6]$ gives:

$$[6]_{\text{tot}}/3K_{(n=0)} = [P-6]^3(1+K_6^3) + 2[P-6]^2[P-2](1+K_6^2K_2) + [P-6][P-2]^2(1+K_6K_2^2).$$

With $K_6 = 1$:

$$= 2[P-6]^3 + 2[P-6]^2[P-2](1+K_2) + [P-6][P-2]^2(1+K_2^2).$$

This equation expresses $[P-2]$ as a function of K_2 and $[P-6]$

From these two equations $[P-2]$ and $[P-6]$ can be numerically calculated for a specific value of K_2 .

From $[P-2]$ and $[P-6]$ a CD-signal is calculated using:

$$\begin{aligned} [P-2_3 \bullet \text{DEB}_6] &= K_{(n=3)}[P-2]^3 \\ [M-2_3 \bullet \text{DEB}_6] &= K_{(n=3)}[M-2]^3 = K_{(n=3)}K_2^3[P-2]^3 = K_2^3[P-2_3 \bullet \text{DEB}_6] \\ [P-2_26 \bullet \text{DEB}_6] &= 3K_{(n=3)}[P-2]^2[P-6] \\ [M-2_26 \bullet \text{DEB}_6] &= 3K_{(n=3)}[M-2]^2[M-6] = 3K_{(n=3)}K_2^2[P-2]^2K_6[P-6] = K_2^2[P-2_26 \bullet \text{DEB}_6] \\ [P-26_2 \bullet \text{DEB}_6] &= 3K_{(n=3)}[P-2][P-6]^2 \\ [M-26_2 \bullet \text{DEB}_6] &= 3K_{(n=3)}[M-2][M-6]^2 = 3K_{(n=3)}K_2[P-2]K_6^2[P-6]^2 = K_2[P-26_2 \bullet \text{DEB}_6] \end{aligned}$$

resulting in:

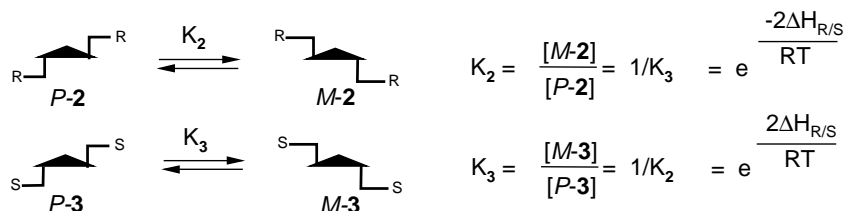
$$\begin{aligned} \Delta\epsilon_{\text{calc.}} &= \text{SF} \frac{([M-2_3 \bullet \text{DEB}_6] - [P-2_3 \bullet \text{DEB}_6] + [M-2_26 \bullet \text{DEB}_6] - [P-2_26 \bullet \text{DEB}_6] + [M-26_2 \bullet \text{DEB}_6] - [P-26_2 \bullet \text{DEB}_6])}{([M-2_3 \bullet \text{DEB}_6] + [P-2_3 \bullet \text{DEB}_6] + [M-2_26 \bullet \text{DEB}_6] + [P-2_26 \bullet \text{DEB}_6] + [M-26_2 \bullet \text{DEB}_6] + [P-26_2 \bullet \text{DEB}_6])} \\ &= \frac{(\text{d.e.}_{6R}[2_3 \bullet \text{DEB}_6] + \text{d.e.}_{4R/2N}[2_26 \bullet \text{DEB}_6] + \text{d.e.}_{2R/4N}[26_2 \bullet \text{DEB}_6])}{(\Sigma[2_{3-n}6_n \bullet \text{DEB}_6])} \times \Delta\epsilon_{(23 \bullet \text{DEB}_6)} \end{aligned}$$

SF: Scaling Factor

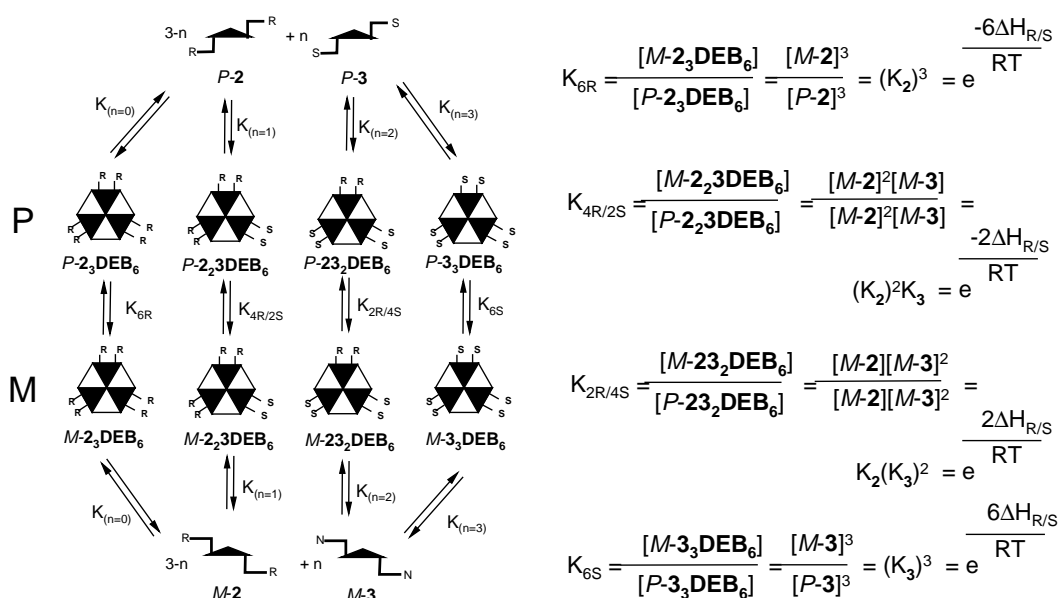
The experimental CD-curve can now be fitted by variation of K_2 using least rms fit analysis. ($[P-2]$ and $[P-6]$ are numerically solved for each K_2 in a subroutine).

Supplementary Material 3.1: Model for Mixing Chiral Assemblies 2_3-DEB_6 and 3_3-DEB_6

1) Define two different conformations M en P for separate building blocks **2** and **3**.



2) Incorporate M/P -conformations without change in energy into the assemblies.



N.B.: cooperativity between the chiral centres is not included in the model !!!

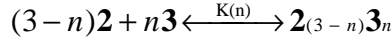
3) Curve fitting procedure (see attachment).

4) Calculate curves and composition of the mixture.

Supplementary Information 3.2:

Curve Fitting Procedure for Mixed Assemblies $2_{3-n}3_n\bullet\text{DEB}_6$ ($n=0-3$).

The equilibrium between assemblies and individual building blocks is described by



$$K(n) = \frac{[2_{(3-n)}3_n]}{[2]^{(3-n)}[3]^n}$$

$3K_{(n=0)}=K_{(n=1)}=K_{(n=2)}=3K_{(n=3)}$, as a result of the three possibilities of forming an assembly with composition **223** (**223**, **232**, and **322**) and **233** (**233**, **323**, and **332**). It is assumed that free components **2** and **3** are not present and therefore $K_{(n)}$ is assigned a value of $1E10$.

Assemblies with *P*-chirality are only formed from combinations of building blocks with *P*-conformation, *e.g.* *P*-**2** and *P*-**3**. The enthalpy of formation is identical for all assemblies (homomeric and heteromeric).

Based on these assumptions the mass balances for $[2_{\text{tot}}]$ and $[3_{\text{tot}}]$ are :

$$[2]_{\text{tot}} = 3[P\text{-}2_3\bullet\text{DEB}_6] + 2[P\text{-}2_23\bullet\text{DEB}_6] + [P\text{-}23_2\bullet\text{DEB}_6] + 3[M\text{-}2_3\bullet\text{DEB}_6] + 2[M\text{-}2_23\bullet\text{DEB}_6] + [M\text{-}23_2\bullet\text{DEB}_6].$$

$$[2]_{\text{tot}} = 3K_{(n=3)}[P\text{-}2]^3 + 2K_{(n=2)}[P\text{-}2]^2[P\text{-}3] + K_{(n=1)}[P\text{-}2][P\text{-}3]^2 + 3K_{(n=3)}[M\text{-}2]^3 + 2K_{(n=2)}[M\text{-}2]^2[M\text{-}3] + K_{(n=1)}[M\text{-}2][M\text{-}3]^2.$$

$$[2]_{\text{tot}} = 3K_{(n=3)}[P\text{-}2]^3 + 6K_{(n=3)}[P\text{-}2]^2[P\text{-}3] + 3K_{(n=3)}[P\text{-}2][P\text{-}3]^2 + 3K_{(n=3)}[M\text{-}2]^3 + 6K_{(n=3)}[M\text{-}2]^2[M\text{-}3] + 3K_{(n=3)}[M\text{-}2][M\text{-}3]^2.$$

$$[2]_{\text{tot}}/3K_{(n=3)} = [P\text{-}2]^3 + 2[P\text{-}2]^2[P\text{-}3] + [P\text{-}2][P\text{-}3]^2 + [M\text{-}2]^3 + 2[M\text{-}2]^2[M\text{-}3] + [M\text{-}2][M\text{-}3]^2.$$

$$[M\text{-}2] = K_2 \times [P\text{-}2] \text{ and } [M\text{-}3] = K_3 \times [P\text{-}3] \text{ gives:}$$

$$[2]_{\text{tot}}/3K_{(n=3)} = [P\text{-}2]^3(1+K_2^3) + 2[P\text{-}2]^2[P\text{-}3](1+K_2^2K_3) + [P\text{-}2][P\text{-}3]^2(1+K_2K_3^2).$$

With $K_3 = 1/K_2$:

$$= [P\text{-}2]^3(1+K_2^3) + 2[P\text{-}2]^2[P\text{-}3](1+K_2) + [P\text{-}2][P\text{-}3]^2(1+1/K_2).$$

This equation expresses $[P\text{-}3]$ as a function of K_2 and $[P\text{-}2]$.

$$[3]_{\text{tot}} = 3[P\text{-}3_3\bullet\text{DEB}_6] + 2[P\text{-}23_2\bullet\text{DEB}_6] + [P\text{-}2_23\bullet\text{DEB}_6] + 3[M\text{-}3_3\bullet\text{DEB}_6] + 2[M\text{-}23_2\bullet\text{DEB}_6] + [M\text{-}2_23\bullet\text{DEB}_6].$$

$$[3]_{\text{tot}} = 3K_{(n=0)}[P\text{-}3]^3 + 2K_{(n=1)}[P\text{-}2][P\text{-}3]^2 + K_{(n=2)}[P\text{-}2]^2[P\text{-}3] +$$

$$3K_{(n=0)}[M-3]^3 + 2K_{(n=1)}[M-2][M-3]^2 + K_{(n=2)}[M-2]^2[M-3].$$

$$[3]_{\text{tot}} = 3K_{(n=0)}[P-3]^3 + 6K_{(n=0)}[P-2][P-3]^2 + 3K_{(n=0)}[P-2]^2[P-3] + 3K_{(n=0)}[M-3]^3 + 6K_{(n=0)}[M-2][M-3]^2 + 3K_{(n=0)}[M-2]^2[M-3].$$

$$[3]_{\text{tot}}/3K_{(n=0)} = [P-3]^3 + 2[P-2][P-3]^2 + [P-2]^2[P-3] + [M-3]^3 + 2[M-2][M-3]^2 + [M-2]^2[M-3].$$

$[M-2] = K_2 \times [P-2]$ and
 $[M-3] = K_3 \times [P-3]$ gives:

$$[3]_{\text{tot}}/3K_{(n=0)} = [P-3]^3(1+K_3^3) + 2[P-3]^2[P-2](1+K_3^2K_2) + [P-3][P-2]^2(1+K_3K_2^2).$$

With $K_3 = 1/K_2$:

$$= [P-3]^3(1+1/K_2^3) + 2[P-3]^2[P-2](1+1/K_2) + [P-3][P-2]^2(1+K_2).$$

This equation expresses $[P-2]$ as a function of K_2 and $[P-3]$.

From these two equations $[P-2]$ and $[P-3]$ can be calculated numerically for a specific value of K_2 .

From $[P-2]$ and $[P-3]$ a CD-signal is calculated using:

$$\begin{aligned} [P-2_3 \bullet \text{DEB}_6] &= K_{(n=3)}[P-2]^3 \\ [M-2_3 \bullet \text{DEB}_6] &= K_{(n=3)}[M-2]^3 = K_{(n=3)}K_2^3[P-2]^3 = K_2^3[P-2_3 \bullet \text{DEB}_6] \\ [P-2_2 3 \bullet \text{DEB}_6] &= 3K_{(n=3)}[P-2]^2[P-3] \\ [M-2_2 3 \bullet \text{DEB}_6] &= 3K_{(n=3)}[M-2]^2[M-3] = 3K_{(n=3)}K_2^2[P-2]^2K_3[P-3] = K_2[P-2_2 3 \bullet \text{DEB}_6] \\ [P-2_3 2 \bullet \text{DEB}_6] &= 3K_{(n=3)}[P-2][P-3]^2 \\ [M-2_3 2 \bullet \text{DEB}_6] &= 3K_{(n=3)}[M-2][M-3]^2 = 3K_{(n=3)}K_2[P-2]K_3^2[P-3]^2 = (1/K_2)[P-2_3 2 \bullet \text{DEB}_6] \\ [P-3_3 \bullet \text{DEB}_6] &= K_{(n=3)}[P-3]^3 \\ [M-3_3 \bullet \text{DEB}_6] &= K_{(n=3)}[M-3]^3 = K_{(n=3)}K_3^3[P-3]^3 = (1/K_2)^3[P-3_3 \bullet \text{DEB}_6] \end{aligned}$$

resulting in:

$$\begin{aligned} \Delta \epsilon_{\text{calc.}} &= \text{SF} ([M-2_3 \bullet \text{DEB}_6] - [P-2_3 \bullet \text{DEB}_6] + [M-2_2 3 \bullet \text{DEB}_6] - [P-2_2 3 \bullet \text{DEB}_6] + \\ &\quad \frac{[M-2_3 2 \bullet \text{DEB}_6] - [P-2_3 2 \bullet \text{DEB}_6] + [M-3_3 \bullet \text{DEB}_6] - [P-3_3 \bullet \text{DEB}_6]}{([M-2_3 \bullet \text{DEB}_6] + [P-2_3 \bullet \text{DEB}_6] + [M-2_2 3 \bullet \text{DEB}_6] + [P-2_2 3 \bullet \text{DEB}_6] + \\ &\quad [M-2_3 2 \bullet \text{DEB}_6] + [P-2_3 2 \bullet \text{DEB}_6] + [M-3_3 \bullet \text{DEB}_6] + [P-3_3 \bullet \text{DEB}_6])} \\ &= \text{SF} \frac{(\text{d.e.}_{6R}[2_3 \bullet \text{DEB}_6] + \text{d.e.}_{4R/2S}[2_2 3 \bullet \text{DEB}_6] + \text{d.e.}_{2R/4S}[2_3 2 \bullet \text{DEB}_6] + \text{d.e.}_{6S}[3_3 \bullet \text{DEB}_6])}{\Sigma_{(n=0-3)} [2_{3-n} 3_n \bullet \text{DEB}_6]} \times \Delta \epsilon_{(23 \bullet \text{DEB}_6)} \end{aligned}$$

SF: Scaling Factor

The experimental CD-curve can now be fitted by variation of K_2 . ($[P-2]$ and $[P-3]$ are numerically solved for each K_2 in a subroutine) using least rms fit analysis.